

Circulating Tumor Necrosis Factor Alpha Concentrations Are Higher in Abdominal Versus Peripheral Obesity

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Fat tissue is a significant source of endogenous tumor necrosis factor alpha (TNF α), the pluripotent cytokine that plays an important role as a mediator of the peripheral insulin resistance found in obesity. The majority of evidence for this role of TNF α is from studies in animal models of obesity. To explore further the role of TNF α in the pathogenesis of obesity-related insulin resistance in humans, we compared plasma levels of TNF α and the other main endocrine cytokine, interleukin-6 (IL-6) both measured by enzyme-linked immunosorbent assay, in 26 obese women (body mass index [BMI] > 30 kg/m²) and 13 female controls (BMI < 26 kg/m²) without a history of recent or active infection. Glucose and insulin levels were measured at 0, 1, and 2 hours after a 75-g oral glucose load. There was no significant difference in plasma TNF α or IL-6 levels between obese and non-obese subjects overall (2.10 ± 0.19 v 1.65 ± 0.18 pg/mL and 2.06 ± 0.29 v 1.50 ± 0.17 pg/mL, respectively). However, TNF α levels were significantly elevated in obese subjects with a 2-hour glucose level more than 140 mg/dL ($n = 8$) compared with the other obese subjects ($n = 18$) and the non-obese controls (2.88 ± 0.46 v 1.75 ± 0.10 and 1.65 ± 0.18 pg/mL, respectively, $P < .01$). Furthermore, the TNF α level correlated significantly with the waist to hip ratio ([WHR] $r = .53$, $P < .01$) and fasting and post-oral glucose tolerance test (OGTT) insulin levels ($r = .47$, $P < .02$), but not with the BMI, and was higher in obese women with a WHR more than 0.90 ($n = 14$) in comparison to those with a WHR less than 0.90 ($n = 12$, 2.47 ± 0.29 v 1.66 ± 0.18 pg/mL, respectively, $P < .03$). The corresponding plasma leptin level was significantly higher in obese women versus the control group (41.6 ± 2.5 v 22.3 ± 2.9 ng/mL, $P < .001$) and was related to the BMI ($r = .60$, $P < .01$) but not to TNF α or the WHR. There were no significant differences in the corresponding IL-6 concentration between groups, and IL-6 did not correlate with TNF α , leptin, BMI, WHR, or insulin levels. In conclusion, circulating TNF α levels are higher in abdominal obesity compared with peripheral obesity, and may contribute to the insulin resistance that more commonly complicates the former pattern of fat distribution.

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TUMOR NECROSIS FACTOR ALPHA (TNF α) is a pluripotent cytokine known primarily for its role in the inflammatory response.¹ TNF α also has important effects on lipid and glucose metabolism.² It has been shown that fat tissue is a significant source of endogenous TNF α production.³ In fact, TNF α expression is elevated in the adipose tissue of most rodent models of obesity and appears to play a critical role as a mediator of the peripheral insulin resistance that characterizes these animals.^{3,4} Recently, it has been demonstrated that TNF α expression is also elevated in human fat and muscle tissue and that it best correlates with the body mass index (BMI) and the degree of hyperinsulinemia.^{5,6} In addition, weight loss caused by dietary treatment in obese subjects results in a significant decrease of TNF α expression in adipose tissue with a parallel improvement of insulin resistance.⁷ This suggests that abnormal production of TNF α may also play an important role in obesity-linked insulin resistance in man. TNF α interferes with insulin action, probably by inhibiting insulin receptor signaling.^{8,9} Thus, TNF α induces serine phosphorylation of insulin receptor substrate-1¹⁰ and converts it to an inhibitor of insulin receptor tyrosine kinase activity. However, the proximal signal-

ing pathways activated by TNF α to inhibit insulin action are largely unknown.

TNF α actions in obesity appear to occur primarily via autocrine and paracrine mechanisms involving adipose and muscle tissue, the principal targets of insulin action.⁴ It is possible that TNF α may also reach targets of insulin action via the circulation, thus having an additional endocrine contribution to the pathogenesis of obesity-related insulin resistance. To explore this possibility, we examined whether TNF α secretion in the circulation is elevated in human obesity and whether it relates to markers of insulin resistance. We also examined whether obesity is associated with elevations in the circulatory level of the other main endocrine cytokine, interleukin-6 (IL-6). Recently, it was shown that IL-6, like TNF α , is also produced and released by human adipose tissue, albeit in smaller amounts that roughly correspond to 10% of total-body tissue release.¹¹ IL-6 increases readily in response to inflammatory and noninflammatory stressors such as trauma or exercise,^{12,13} and has important endocrine and metabolic effects that may influence glucose homeostasis.^{14,15} By measuring circulating IL-6 levels, we wished to control for nonadipose sources of TNF α (mainly inflammatory) and for the possibility that IL-6, independently of TNF α , also may contribute to the metabolic abnormalities found in human obesity.

SUBJECTS AND METHODS

We studied 26 obese women with a BMI more than 30 (calculated as the weight in kilograms divided by the square of the height in meters) and 13 age-matched non-obese controls with a BMI less than 26 without a history of recent or active infection or inflammatory disease. None of the subjects were on medications that affect glucose homeostasis or sympathetic nervous system activity. All subjects underwent a 75-g oral glucose tolerance test (OGTT) with measurement of plasma glucose and insulin levels at 0, 1, and 2 hours after the glucose load. A blood

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Submitted February 18, 1999; accepted April 26, 1999.

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0026-0495/99/4810-0022\$10.00/0

glucose level more than 140 mg/dL at 2 hours after the OGTT was used as the standard for impaired glucose tolerance. Venous blood samples were collected after a 10-hour fast, the plasma was separated immediately, and aliquots were frozen at -40°C until needed for assays. Basal plasma levels of TNF α and IL-6 were measured in duplicate in a single assay by high-sensitivity enzyme-linked immunosorbent assay kits (Quantikine; R & D Systems, Minneapolis, MN). The assay sensitivity was 0.3 pg/mL for TNF α and 0.2 pg/mL for IL-6. The intraassay coefficient of variation (CV) was less than 5% for both cytokine assays. Serum leptin was determined by a specific commercial radioimmunoassay (RIA) kit (Linco Research, St. Charles, MO). The assay sensitivity was 1.0 ng/mL and the intraassay CV was less than 5%. Serum insulin was determined by RIA and glucose by an in-house assay (glucose oxidase method).

Data are expressed as the mean \pm SE. Comparisons between groups were made by the Mann-Whitney test, and the relationship between individual variables was tested by Spearman's rank correlation analysis.

RESULTS

Clinical characteristics of the obese and control patients are summarized in Table 1. There were no overall significant differences between obese and control subjects for either plasma TNF α (2.10 ± 0.19 v 1.65 ± 0.18 pg/mL) or plasma IL-6 (2.06 ± 0.29 v 1.50 ± 0.17 pg/mL), although the levels of both cytokines tended to be higher in obese women.

However, after grouping obese patients on the basis of the OGTT response (2-hour post-OGTT glucose >140 mg/dL, $n = 8$; 2-hour post-OGTT glucose <140 mg/dL, $n = 18$), TNF α levels were significantly higher ($P < .01$) in the former (2.88 ± 0.46 pg/mL) compared with the latter (1.75 ± 0.10 pg/mL) and with non-obese controls (1.65 ± 0.18 pg/mL). The corresponding IL-6 levels showed no significant differences between the three groups (Fig 1).

Similarly, when obese patients were grouped on the basis of the WHR as subjects with abdominal fat distribution (WHR >0.90 , $n = 14$) and subjects with peripheral fat distribution (WHR <0.90 , $n = 12$), plasma TNF α levels were significantly higher in the former group (2.47 ± 0.29 pg/mL) compared with the latter (1.66 ± 0.13 pg/mL, $P < .03$). In contrast, IL-6 levels remained comparable between the groups (1.90 ± 0.21 v 2.25 ± 0.44 pg/mL) (Fig 2).

Consistent with these data, the TNF α levels correlated

Table 1. Clinical Characteristics of the Obese and Control (lean) Subjects

Characteristic	Obese			
	Lean (n = 13)	Total Group (n = 26)	IGT (n = 8)	NGT (n = 18)
Age (yr)	46.5 ± 3.1	42.1 ± 2.6	49.1 ± 4.6	39.5 ± 2.9
BMI (kg/m^2)	24.4 ± 1.6	$39.5 \pm 1.3^{\S}$	$39.3 \pm 1.5^{\S}$	$39.9 \pm 1.9^{\S}$
WHR	0.85 ± 0.03	$0.90 \pm 0.02^{\ddagger}$	$0.99 \pm 0.01^{\dagger}$	0.86 ± 0.02
Glucose (mg/dL)				
Basal	93 ± 3	91 ± 4	107 ± 8	83 ± 3
2 h post-OGTT	119 ± 4	$135 \pm 9^{\ddagger}$	$177 \pm 12^{\dagger}$	110 ± 4
Insulin (mIU/mL)				
Basal	19 ± 4	$28 \pm 4^{\ddagger}$	$31 \pm 3^*$	26 ± 4
2 h post-OGTT	109 ± 26	139 ± 22	$199 \pm 30^*$	108 ± 25

Abbreviations: IGT, impaired glucose tolerance; NGT, normal glucose tolerance.

* $P < .05$, $^{\dagger}P < .01$ v lean and obese NGT.

$^{\ddagger}P < .05$, $^{\S}P < .01$ v lean.

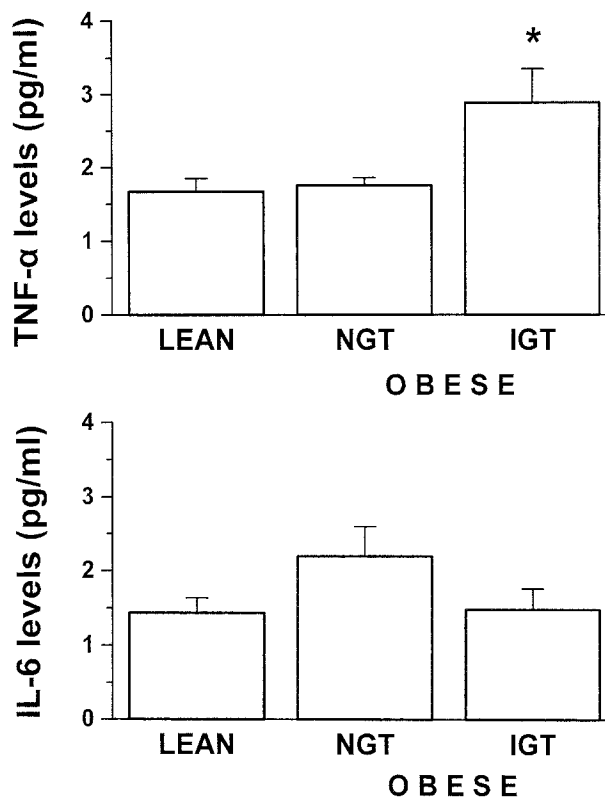


Fig 1. Plasma TNF α levels were significantly higher ($P < .01$) in obese women with impaired glucose tolerance (IGT) compared with obese women with normal glucose tolerance (NGT) and lean controls. The corresponding plasma IL-6 levels were not significantly different between groups.

significantly with the WHR ($r = .53$, $P < .01$) and with fasting, post-OGTT, and integrated (area under the curve [AUC]) serum insulin levels ($r = .47$, $P < .02$), as well as the ratio for the AUC of insulin to the AUC of glucose ($P < .05$), in obese patients (Fig 3). In contrast, TNF α did not correlate with the BMI ($r = .30$, $P = .12$) or with leptin levels in these patients, the latter being clearly elevated compared with the value in the lean controls (41.6 ± 2.5 v 22.3 ± 2.9 ng/mL, $P < .001$) directly proportionately to the BMI ($r = .60$, $P < .01$).

There were no differences in TNF α or leptin levels between premenopausal ($n = 18$) and postmenopausal ($n = 8$) obese women (2.01 ± 0.23 pg/mL and 40.8 ± 3.6 ng/mL v 2.07 ± 0.20 pg/mL and 42.4 ± 5.3 ng/mL, respectively).

DISCUSSION

This study shows that circulating TNF α levels are clearly elevated in obese women with abdominal obesity, most of whom also have some degree of insulin resistance and/or glucose intolerance. In contrast, TNF α levels are not elevated in women with peripheral obesity. This differential relationship between TNF α and the two patterns of fat distribution is also reflected in the significant positive relationship of TNF α to the WHR, a good, albeit indirect, marker of visceral fat mass, but not to the BMI, a marker of total adiposity that cannot account for fat distribution.

The circulating levels of TNF α found in the present study

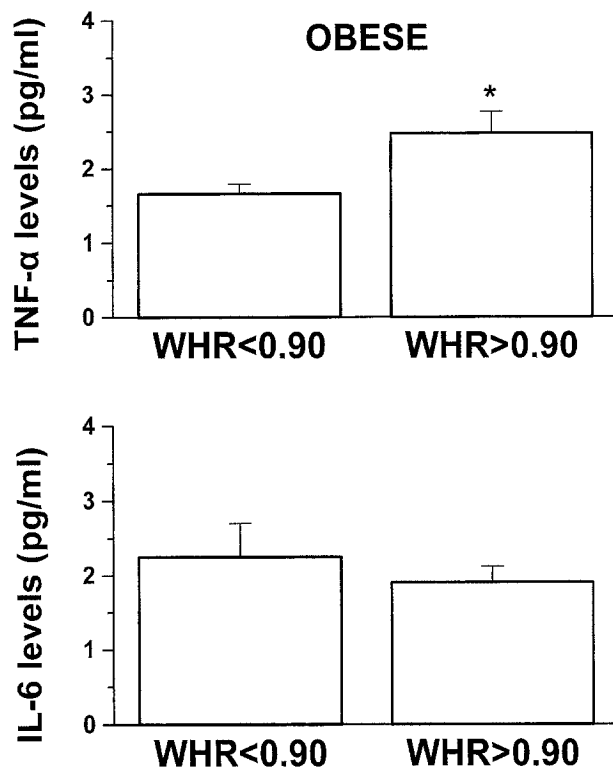


Fig 2. TNF α , but not IL-6, was higher in obese women with a WHR > 0.90 versus a WHR < 0.90.

were relatively low, and it could be argued that they may not have significant biological activity on insulin target tissues. However, it is possible that circulating TNF α acts synergistically with locally produced TNF α in fat and skeletal muscle. The bioavailability and/or action of circulating TNF α in obesity might further depend on the circulating amount of soluble TNF α receptors, which can either inhibit or enhance the activity of TNF α .^{16,17} Moreover, it is believed that in the absence of an inflammatory process, circulating TNF α levels reflect TNF α message expression and protein synthesis in adipose tissue and its constitutive secretion.⁴

The potential contribution of elevations in circulating TNF α to the development or deterioration of insulin resistance and glucose intolerance is supported by the fact that TNF α levels correlated significantly with both the 2-hour post-OGTT glucose level and the fasting, post-OGTT, and AUC insulin levels in our obese subjects. Such a role for circulating TNF α is also suggested by the reported significant decrease of TNF α in obese type 2 diabetic patients after weight loss with dietary treatment and exercise in parallel with an improvement in insulin sensitivity (decrease in insulin).¹⁸ Interestingly, in the same study, consistent with our findings, basal TNF α levels correlated with the visceral fat area and not with the subcutaneous fat area (both measured by computed tomography) or BMI.¹⁸ In another study, serum TNF α bioactivity was increased in obese and type 2 diabetic patients and correlated positively with basal serum C-peptide levels.¹⁹ Also, Dandona et al²⁰ have shown that serum TNF α decreases with weight loss in obese patients.

In addition to the direct inhibitory effect of TNF α on insulin signaling, there is evidence that elevations of circulating TNF α

may also contribute to the increase in fibrinogen and plasminogen activator inhibitor-1 production and to the lipid abnormalities (high total cholesterol and triglycerides and low high-density lipoprotein cholesterol) that, together with hyperinsulinemia, are important components of the metabolic syndrome.^{21,22}

It is of note that TNF α did not correlate with serum leptin, despite *in vitro* and *in vivo* evidence that TNF α may contribute to obesity-related hyperleptinemia by stimulating leptin release from adipocytes.^{23,24} This suggests that the regulation of leptin production is largely under the control of different factors.

Unlike TNF α , circulating IL-6 did not differ between obese and lean individuals and did not relate to fat distribution or to indices of insulin resistance. This suggests that in the absence of infection or inflammation, circulating IL-6 does not have an appreciable contribution to obesity-related insulin resistance, and TNF α and IL-6 expression may be subject to different regulatory mechanisms. Further, the lack of correlation between IL-6 and the presence or degree of obesity in our patients suggests that the recently reported increases in serum IL-6 in obese sleep-apneic patients²⁵ are most likely a correlate of the sleep apnea and related hypoxia rather than obesity *per se*.

In conclusion, circulating TNF α , but not IL-6, is higher in abdominal compared with peripheral obesity and may contribute to the insulin resistance that more commonly complicates the former pattern of fat distribution.

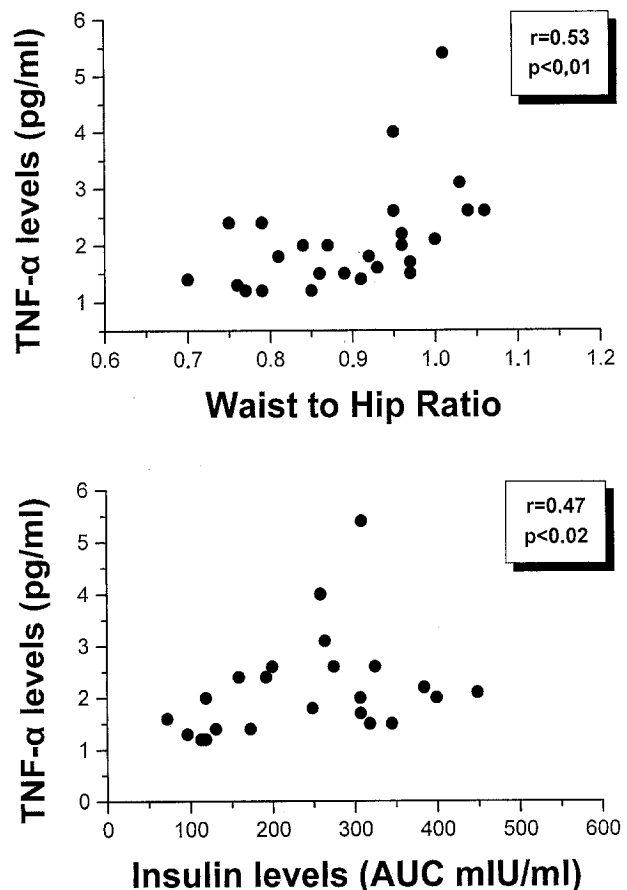


Fig 3. Plasma TNF α correlated significantly with the WHR and the integrated insulin level (AUC) in obese patients.

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